

March 4, 1954

Interpretation of origin and behavior of the controlling systems.

I. Review of previous discussion.

1. Purpose was to show that systems present that control genic action in manner not quite like that of Ds - Ac or that of a_1 - Dt, (which are much alike).

2. These other systems, have basic pattern however that is similar -- control of genic action; evidence suggests that segregations of controlling units occur. Meiotic segregations obtained and linkages noted. Results of somatic segregations similar to those of meiotic segregations.

II. Controlling systems revealed by the b.f.b. cycle. Concluded that they do not arise denovo but are present in the normal nucleus -- function in some precise way during the development of the organism.

a). Somatic segregations occur, changing the constitutions of the sister nuclei with respect to the controlling unit.

b). These changes result in subsequent controls of the action of specific genes.

c). For the genes to become active, some change must occur at the locus of the gene.

d). Some mechanism must be present in the nucleus that initiates these changes in nuclear organization and constitution.

e). Believe this mechanism associated with the heterochromatic elements of the chromosomes -- primary initiators of the changes that subsequently occur.

f). The pattern controlling genes, the "switch" genes, etc., may be expressions of their action.

III. The recent studies of others showing the mechanism outlined in this series of discussions:

1. Work of Brink and students on the var. P locus in maize: Have come to same conclusions regarding the controls of the patterns of variegation expressed. Have found the transposing units. Have found the twin sectors of the types described in this series and have investigated the progeny derived from them. Confirms the story here presented.

2. Peterson, in maize, has found system controlling variegation that also appears to follow the main conditions here discussed -- the controls of genic action; transpositions of the controlling unit comparable, in many respects to Ac but not Ac.

3. The studies of the "Hi" mutation factor in Drosophila by Ives and Hinton -- suggests controlled changes at certain gene loci and controlled types of chromosomal alterations that occur at specific loci.

4. Somatic crossing-over as observed by Stern. Some cases strongly suggest a controlled system in operation here to give precise types of exchanges between certain chromosome components.

5. Many suggestive cases in literature. The cytological cases are most instructive in this respect. Review will be given of representative cases

IV. According to interpretation given -- that the effects described represent those normally occurring in development -- we must account for their appearance in the stocks used here.

1. If the processes described of control of genic action occur in standard strains, their presence not readily revealed. This is because they are active at particular times, controlled in this respect, and therefore not observed as specific units unless they are called modifiers.

2. If the controls not regulated, a chaotic situation would occur, regarding gene action, the regulation of development, the maintenance of species, etc.

3. The pattern controlling genes -- look very much like the systems outlined

a). The patterns in flowers: strikingly controlled in many cases.

b). The patterns of pigmented areas in the Lady Beetle -- again, a striking case of control of when the gene for pigment formation, or involved in it, is effective.

c) switch gene -

4. If one of these systems of control becomes "out-of phase" -- starts working at a different time in development, the results would be apparent. Altered times of gene action would be noted. Variegation would result.

5. The mechanism of change, as here outlined in the discussions, would make it clear that once such a system becomes "out-of-phase", it would continue to be so, and would result in the appearance of new changes in genic action, elsewhere, which then would be put "out-of-phase".

V. The interpretation described would require that in the normal development of an organism, somatic segregations should occur. The nuclei should not all be alike. Again, changes at particular gene loci should occur at specific types, leading to nuclei with different potentialities for genic action.

What about somatic segregations?

1. What do we know about somatic segregations? About differences in genic action in different nuclei of an organism? Is there any evidence to support the above interpretations?

2. There is a vast wealth of information in support of the ideas given. These are mainly in the area of investigation known as "cytology" and much of it is purely descriptive. Many of the papers in this field are not examined by geneticists because of their highly technical nature. Most are not seen at all by biologists in other fields. The observations, however, are of vast significance. The field has only been tapped and a wide opportunity is present for experimental investigations.

3. A brief review will be given of the nature of this evidence, with examples selected for illustrations. Will cover the following topics:

- mp {
- (1) Differences in nuclei of organism with respect to sets present.
 - (2) Differences in nuclei of organisms with respect to particular chromosomes present.
 - (3) Differences with respect to component parts of the chromosome itself:
 - (a) Large segments to
 - (b) the minutest segments known -- the bands in the salivary gland type chromosome.

VI. Differences in nuclei of an organism with regard to the number of sets of chromosomes present:

1. Zygote -- starts out with two sets of chromosomes, one from male and one from female. During development, changes occur in the number of sets of chromosomes within the nuclei. These are controlled changes: certain types of cells having certain specific types of changes in sets. The set may increase -- poly ploid nuclei; or the set may be reduced -- to haploid nucle. These changes occur at specific times during the development of the organism. Also, different methods of change in set number occur.

The increases in numbers of sets: By means of endomitosis -- no mitotic division figures. Various methods of accomplishing this endomitosis. Fusions

a). The degrees of increase: From tetraploid to 2,000 ploid.

b). The type of appearance of chromosomes: Polytene type - salivaries or diffuse type -- frequent; Or, combination of both.

The decreases in number of sets: Some interesting examples:

a). Tail of Rana (frog). Epidermis is diploid (Groom, 1957)
Mesenchyme is haploid (Ludlow, 1953)

b). Sea Urchin: A "reduction division" in certain cells early in cleavage stage: 16 cell stage -- 4 micromeres present. These are 2n. In division of micromeres, reduction of chromosome number occurs to haploid. These haploid cells become skeletal components of organism.

b). Increase of sets followed by decrease of sets: Culex. Very well examined case to show the mechanism involved. Reduction occurs during metamorphosis period.

VII. Differences in nuclei with regard to certain chromosomes of the complement: Loss of chromosomes at specific divisions: ~~translocation studies~~
Non-disjunctions of certain chromosomes in certain divisions; Other methods of accomplishing the same.

1. Best worked case that of Sciara: Outline on board.

Loss of chromosomes at mitosis

Non-disjunction of particular chromosome at certain division

Passage of chromosome through nuclear membrane -- highly controlled.

The experiments that show which part of chromosome is responsible for the phenomena.

When is this effect impressed on centromeres?

What mechanism makes the impression so precise?

2. The B-type chromosomes in plants. Very highly controlled non-disjunction of these chromosomes. Occur at specific times and with specific consequences.

a). Example of this behavior -- the B chromosome in maize:
Diagram on board.

Control exerted at centromere region of B-chromosome, as in the X chromosomes in Sciara. The translocation studies showing this.

3. Ascaris - loss of parts of chromosomes - translocation studies.

VIII. Differences in behavior of sets of chromosomes within same nucleus:

Ch. no. condensed = 28.

White . Monarthropalpus. Spermatogenesis:



→

Sperm receives chromosomes in condensed set.

IX. Differences in appearance of different chromosomes of a set in different stages.

- a). The small chromosome in salivary glands of White.
" " " " "elland
- b). The alternation in condensation of particular chromosomes:



Type 1

Type 2

X. Differences expressed by individual parts of chromosomes in different cells of an organism:

- 1). The chromocenter regions -- heterochromatin in Drosophila -- too well known to comment on.

- 2). Condensed versus diffuse condition of segment of a chromosome:



Type 1

Type 2

- 3). Most exciting recent case -- that of Pavan on Rhyndhisclara -- two different species studied.

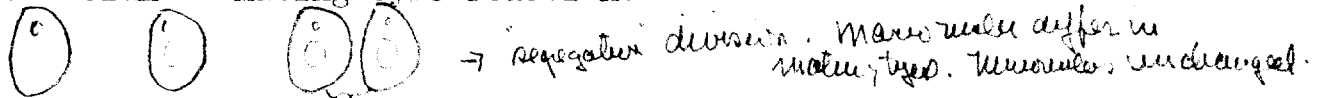
- a). All eggs laid at one time -- within 45 mins.
b). Nearly all larvae of one sex
c). All develop at same rate - 42 days to pupation. Common puparium formed
d). The changes occurring to specific bands at certain times in development in the different tissues; change at particular band specific for the tissue involved.
e). One type: at certain time, increase in D.N.A. and swelling of particular band; Separation of components of the polytene bands -- Large Puff region formed. At certain stage, shrinkage of puff region. Return to dense band, but now had very much wider than before the process started. Increase in D.N.A. remains. Interpretation: Multiplication of genes of this band, specifically. Differential behavior.

XI. Conclusions on nuclear compositions in organisms: Wide differences among the cells of an organism with regard to the nuclear composition. Differs with respect to sets, parts of set, parts of chromosomes, even parts of so small as a band in the salivary type chromosome.

Mechanisms : Differential behavior at certain mitoses: Losses of certain chromosomes; Non-disjunctions of controlled type at certain divisions. Passage of chromosome through nuclear membrane.

XII. Differential nuclei resulting from specific divisions: on genic level.

1. Paramoecium -- mating type reaction:



2. Tetrahymena -- mating type reaction: Same as paramoecium except the division is not as precise -- first or second division of macronucleus .

3. Case in Glomerella where mating type "mutation" occurs at specific time in development of ascogenous hyphae.

XIII. The effects of species crosses: Often quite bizaar.

1. Changes in the chromosome complement: Various polyploid conditions, reductions, occur in tissues that do not show such reactions in the individual species involved.

2. Case of Metz in Sciara: Cross of S. x S.

The mosaic salivary gland:

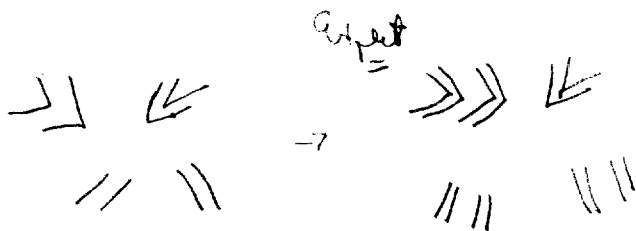
3. Species cross in Nicotiana. A. x B. Backcrossed to A

A.A B complement: B chromosomes eliminated somatically.

3. Altered genic action often noted; Variegation appears.

XIV. The constitution of differentiated nuclei -- as shown when these cells forced to divide.

a). Treatment of root -- chemical (indol-acetic acid). Does not affect behavior of chromosomes in the nuclei that regularly divide. Forces nuclei that are fully differentiated to divide. They show a variety of chromosomal aberrations. One type observed in cell that had undergone endomitotic division:



The effects observed are very much like those produced by Ds.

XV. Ds type behavior in other organisms than maize:

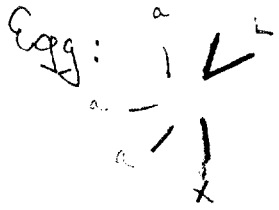
1. Metz - Sciara. Certain chromosome, certain band region -- break seen in all salivary gland cells. Showed that this was genetically controlled.
2. Hyacinthus - Darlington. Break in a particular chromosome late in development of the sporocytes. Genetic control
3. Vicia - Levan. With certain treatment, break occurs at the nucleolus organizer quite predictably.
4. Dreyfus -- break in arm of particular chromosome at certain division in the spermatogenesis of .
5. Catcheside - Oenothera. His case of position effect following translocation: Evidence much like that found for Ds. Chromosome breaks; chromosome duplications and deficiencies of certain region of a chromosome in the translocation complex.

XVI. Conclusions:

- 1.

on locus

Sperm -



x Sperm



Zygote



First 4 cleavages = ordinary mitosis

Separation of germ line + soma



Soma

germline

1 1 1 1 1 1

5th or 6th div

1 1 1 1 1 1

elimination of 1 V not known

To become ♀

To become ♂

1 1 1 1 1 1

1 1 1 1 1 1

=

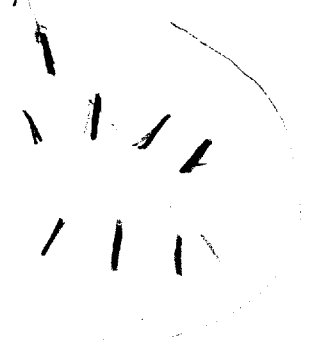
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1 1 1 1 1 1

1 1 1 1 1 1

elimination of 1 x chr.

passes through nuclear membrane

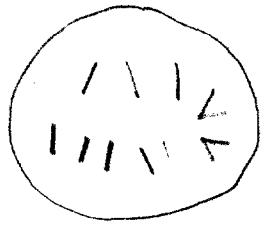


1 x = 1 ♀ : 1 ♂

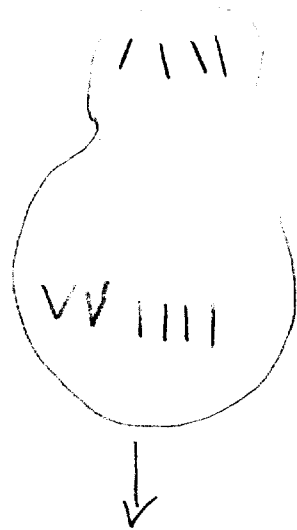
1 x = 1 ♀

The spermatogonium - meiotic divisions.

Prophase: no synapses



Anaphase I



→ Bud; complement form of autosomes and x
Does not develop sperm

Anaphase II



→ non functional cell.



→ sperm nucleus

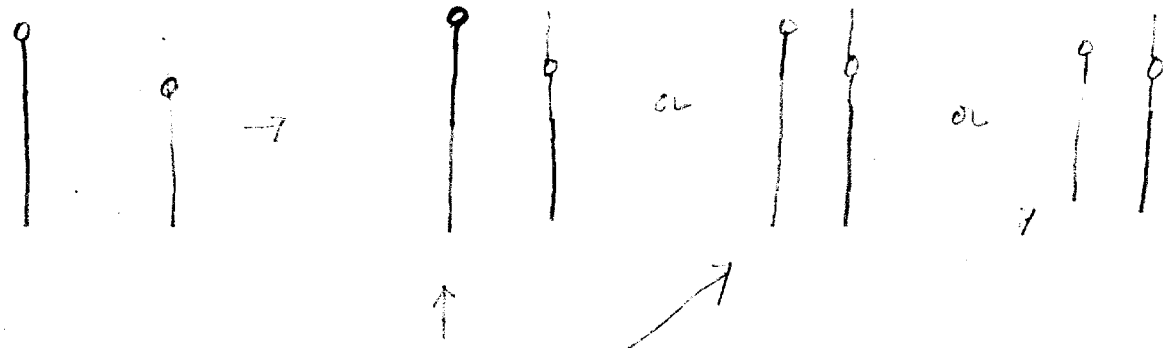
1 set of autosomes from mother

2 x chromosomes from mother

2 identical chromosomes.

Sereno - The translocation studies of Acorns.

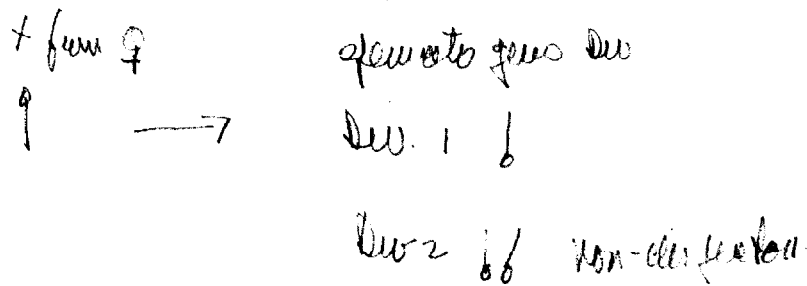
1. Sereno x - autosome translocation



- 1) non-disjunction in female
- 2) Stimulated in cleavage divisions in zona; through nuclear membrane in germ line

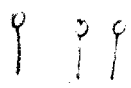
2. The control exerted on by the ^{in control} cellular of x chromosome. not a component of the chromatin in the chromatin as a whole.

3. Alteration of controls in alternate generations: not the effect of particular centromeres which chromatin differentiates. It becomes differentiated at some time.



Zone of male

germ line of one chromosome



chromosomes



chromosomes

chromosomes


non-disjunction

♀♀

4. Diffusion must be imposed on the homologues at some time
during development and this at the entrance of a structure which
prepares it for the eventuality.


The B-type chromosomes in maize -

1. Nature of B-type chr. Extra chromo; not like regular normal complement. Found in various strains of maize

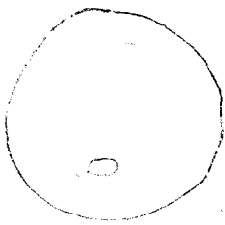
2. The internal morphology of the B-type chr. 

3. Behaviour of B-type in mitosis -

Divides in each mitosis along with regular set of chroms.

Occupies a particular position on plate  at edge of plate.

4. Behaviour in pollen grain divisions: Differential divisions



generates all

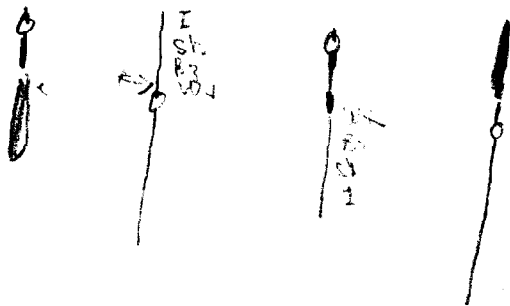
no B

no B

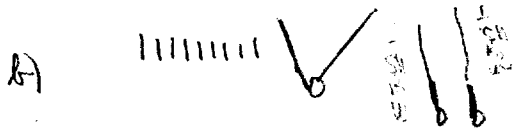
no B: after division;

5. 2 species: one with no B; 1 with 2 Bs.

6. Translocation between A + B shows:



7. The differential division in pollen.



8. Final way of sperm in cross wall Cshyex:

$$\begin{cases} \text{Endosperm with a} = \text{Cshyex in place to} \\ \text{Zygote " b) = Rep. of ISH R, Lx} \end{cases}$$

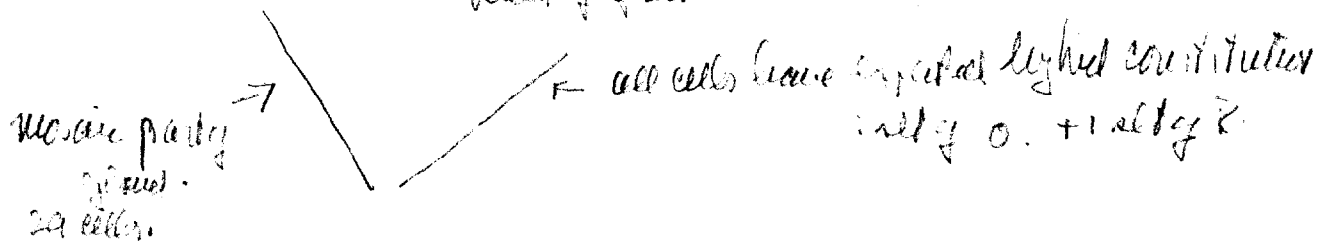
$$\text{Endosperm with b) = ISH R, Lx}$$

$$\text{Zygote " a) = Place = } \frac{b}{a}$$

9. Again, region of cell lines responsible for the non-disjunction in the pollen division.

The main salivary gland in *Parva species* even. - 16×8

1. Ocellars & \times S. Regulatori $\delta 7$.
2. Salivary band structure of each recognizable.
3. The hybrid - one individual -
2 components of gland



16 cells = expected chromosomal constitution.

1 cell = " " " " + 1 extra chromosome from O.

12 cells = all have 2n but all are ocellars clones.

4. Dist. distribution of 12 cells = not contiguous. Appears as if some events occurred in making different cells.

1. Group of ocellars all Regulatori = 5:1 complement

2. Interposition test: is this an example of out-of-phase action usually occurring at mitosis? Expression of chromosomal complement of O?

I. In studies presented, have shown that units appear in the nucleus that differ in quite recognizable way from those usually prepared for the gene:

1. We have considered the gene as an individual entity, occupying a specific locus in a specific chromatin

2. We have considered that this gene operates in a particular way - it is operable, at some level, for the organization of nuclear components which influence the ensuing biochemical reactions in the cell.

3. We have considered that each gene functions to this extent in its own particular way

4. We have considered, ^{in the past} that during the mitotic cycle, each gene is replicated, usually accurately, so that two sister chromatids are able in their joint genetic constitution.

5. We have not been so precise in our thinking of the changes that occur at the gene level when a mutation occurs.

a) A few geneticists have desired to consider a mutation as a change in structure of the gene entity in such a way that it subsequently functions to produce a modified molecular component which then enters into the biochemical sequence. This would be a real gene change.

6). The studies in *Drosophila* of position effects, first observed and defined by Sturtevant, opened up a wide field of research. These studies gained from these studies, a ~~complex~~ ^{mutational or a plasmid} series of modifications of this view of chromosomal structure, the gene entity, and superimposed evidence in projection of the nature of change at the gene level that is responsible for the observed mutation.

a). The position of gene entities along the chromosome remains. We know that if some alterations occur ^{in a gene} at a particular locus in the chromosome, a particular genetic type of alterations could occur.

a). Many regions in the chromosome, however, where we change of a mutation type can be obtained. Instead, changes passed between: alterations of many regions result in variability. This however a big blank in our knowledge of gene composition of the chromosome.

II. In the many studies, the evidence, a second system of ^{chromosomal} control is superimposed on the ^{non-recessive} recessive system of Sturtevant's genes. This system is composed of ^{recessive, sporadic} genetic units, but these units are not stationary. They can move from one location to another.

2. These units differ from the stationary units, in that they can control the action of the stationary units.

3. These movable units have specificity. There are different types of these units, forming specific systems, such as the D-Ac system, the "a" unit - Dt system, the "pg" - En unit system. The action of a gene, in the case of the various numbered genes, reflects the behavior and specificity of the particular controller system in relation to the action of the stationary unit - Examples of A₁:-

1) System "a" - Dt

2) " D - Ac

3) " a^{m-1}

4) " a^{m-2}

Each system differs in its mechanism of control of the gene expression of the stationary A₁ unit.

II. We outlined the evidence of the kinetic changes that occur to chromatin when a element of the complement is undergoing the b-f-b cycle and discussed that certain chromatin components - the cellulose, the histone & the nucleosomes of DNA, were being altered by this process.

1. We have indicated that in plants that underwent this cycle, the systems controlling gene action were made evident.

2. We outlined one test that supports the assumption that the b-f-b cycle could expose more units - the a₁ / a₁ b-f-b cycle stage.

to induce mutation at a.

14

3) we have shown that new unstable genes can arise in plants having a stable genome once that of the system operating at this new locus is the same as that present in the plant -- the numerous cases of origin of the controlled instability when both are present.

4) we have outlined our knowledge of the events that occur at the locus and come to the conclusion in the D. H. case, that the ^{change in phenotypic expression you get to a gene in the genome} gene ~~is not~~ ^{is} ~~mutability~~ but that the changes really were associated with changes of the genome which controls the degree & type of behavior of the gene.

5) we have given the evidence which suggests that the type of change is a result of some physical alteration in the chromatin material of or at the locus.

III. we are left with the problem of accounting for the appearance of these ^{gene} controlling systems: where do they come from? Why and how appear in these plants that underwent the J.B. cycle?

IV. my speculations on them run somewhat as follows:

1. These gene controlling systems are extremely present in higher
2. they operate in "normal" nuclei just as they do in our ^{and other plants} unstable strains. Controlling the action of some genes in the same manner & under the same procedure.

3. They were made broader to us because their limit of operation was thrown "out-of-phase" during the developmental cycle: They are acting at times not associated with the normal cycles.

4. The mechanism responsible for this is associated with the elements involved in the heterochromatin elements of the chromatin.

5. Changes in the quantities and ^{mutual} associations of the heterochromatin elements in some manner initiate changes in the timing of action of the mobile gene controlling units. These mobile units are in the context of the material of the heterochromatin elements but changes in these elements affect the action of the mobile units, putting them "out-of-phase" in the period of action; Once out-of-phase, they remain that way, allowing their moving operation to be discovered.

6. On this view, we are not creating any new phenomena, nor have we introduced a new condition, such as a virus, that would be responsible for the results obtained.

7. Do not patterns in development suggest the presence of new units? Such simple pattern as the position, the size, + the whereabouts phenomenon of the pigmented spots on the of the Lady Beetle? Why does pigment appear only at certain places + at certain times in the formed individuals which reflects the whole growth process function in development? What about the Rhesus in monkeys that show such

precise controls in the development + intensity of pigment in
different tissues and parts of tissues in an organism? Why are there
so many cells of the same ^{local} showing specific action with
respect to these differences + ^{intensity} ^{direction} ^{range} within the same place?
Could there be reflection of the action of controlling units? ^{As} ^{and at different}
changes occur during development at specific times + in specific cells that
control when + how much pigment will be placed in the concerned
cells?

8. If the assumptions made above were true, then, the making
of an organism should not be ^{+ controlled} but should differ from
no other in very precise way.

9. The evidence for difference in the genetic control ^{and} ^{level}
and the evidence of how this ^{is} ^{not} ^{to} ^{adequately}
Consider this evidence would require a whole series of actions.

10. I intend, nevertheless, to outline some of this evidence,
in order to indicate the ^{that are being} ^{assess} ^{range} of differences and the nature of their
origins, when this is known. Only examples of some types, by no
means all types, can be given at this time.

11. These range from differences in ^{involving} number of sets of
chromosomes involving known genes, on both a morphological and physiological
levels.

changes in chromosomal constitution at certain times during development.

I. Polyploidy - "normal" individuals

1. widespread phenomenon. Certain cells of organisms are polyploid -
Degree varies from 1n to 2,000n.
Disruption of chromosomal number levels - ① polyploidy, ② induced strains, ③ combinations.
2. modes of producing polyploidy - various: Colchicine; nuclear fusion; suppression & surface separation.
3. Expression - a) Polyploidy occurs in all taxa - very well known.
b) Polyploid cells in plants - "clonal" - decreasing
certain cells in a tissue
c) Same applies to animals.
4. Not only increase in chromosomal constitution decreasing post-embryonic stages, but also decrease in chromosomal constitution.
a) Most familiar = meristic division
b) less " = Reduction in early developmental stages =
Frog (Green, 1953) - in tail of frog, the meristematic cells have reduced chromosomal number. These of course have the 2n number.
See Urdahn - (Lindahl, 1953) - 16 cell stage, 4 meristematic cells at veg. pole. all with 2n = 36.
Diminishing number of meristematic cells =
cells found have 18 chs;
(some still at 36 chs).

II. Changes in constituents during development - specific elements.

1. Best case became best studies - Pavan.

Outline: The origin of stem cells

" " " " egg "

The focus = Stem

a) Long lived cells

b) " " extra cells - which are

losing in germ line; extra cells -

the region controlling loss = centromeres.

2. Ottos - Cerebrum - White -

3. Differences within nuclei having, polytropic elements:

a). Whole elements - White, Muller, Pavan.

b). Parts of elements at certain times during development -
Beeman, Michaelis, Pavan.

① The Pavan studies: Outline.

c. Schultz - The Y-chromosome cells.

III. The changes in inheritance of nuclei at specific times -

a) the B-type cells = many of them.

a) the component responsible in meiosis. A-B translocation. The
certain region or cluster = on in Pavan.

VI. Differences in behavior of clonings within a nucleus:

a). One set of clones = all considered = strain very different from the other as

b). "clones" = " " at one stage; different, like other clones at another stage.

c). One region or band of a set - differences at different stages of development with set clones of moles.

VII. Conclusions:

1. The ^{different} nuclei of an organism can vary in their gene ^{in expression} variations in a very wide range of possible ways: From differences at specific loci to differences in whole clones, to differences in whole sets:-
 Promoter clones; clones in frequency of clones in families; clones in chromosomal sets; differences in structure - at all levels from a band to a whole set of clones.

2. In the light of this knowledge, the origin and behavior of mutable genes, as outlined, is not a fantastic proposition, but a working hypothesis with much latitude for future experimentation to design, modify and refine our appreciation of the various systems within the nucleus which operate to control the expression of genes during the development process.

Hanney, D.L. + P.A. Caughey

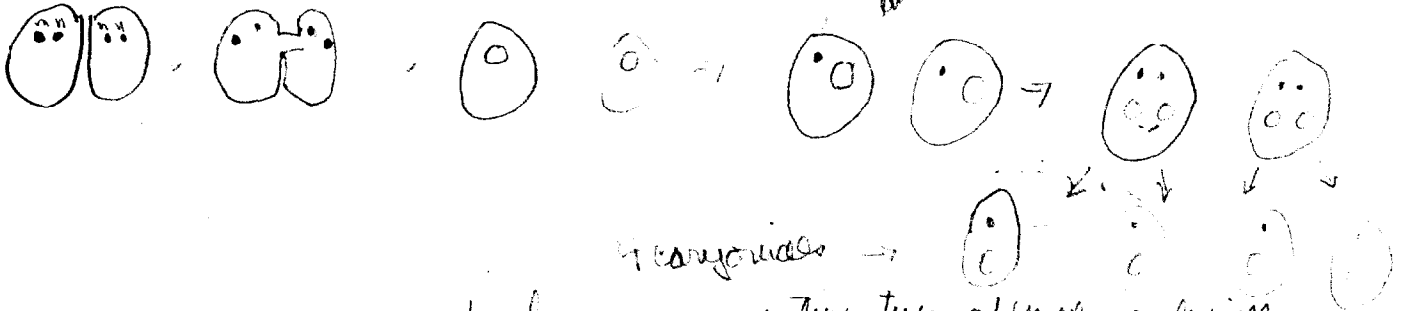
Mating type determination in Tetrahymena pyriformis.

Proc. Nat. Acad. Sci. 39: 113-119

1. m. Paramecium bursaria, multiple mating types occur.
 " aurelia, no more than 2 mating types found within a variety (given 5:10, 1:1000).
2. Tetra. pyriformis: wide variety of forms.
 - 1) amiconucleate, no known sexual form.
 - 2) "selfing" lines
 - 3) crossing clones. This type used: Woods Hole 14, Woods Hole 6: mating type II

3. Lifecycle:

Macros,
1 nucleus divides
= 2 nuclei
divides in 2



4. Among 500 F₁, F₂ + F₃ and backcross, 7 mating types appeared, each will mate with another type but not with itself.
5. Selfing clones also appear. These give as many as 4 mating types within clone, but individual isolates give rise to pure mating types.
6. In P. aurelia (Sommer) mating type within individuals of conjugates is usually stable but mating types produced by sisters (sister conjugates) often differ. Mating type "regulates" at 1st division of macronucleus. Time of change.
7. The Tetrahymena results: Separated cells from 4 conjugates up to those produced by conjugates =



all possible comparisons of mating types in related clones tested. Results of 3 exp. shown in table 2.

Table 2 of paper

Cross	sister sub-conjugates		sister conjugates		co-conjugant conjugates	
	alike	different	alike	different	alike	different
1	7	6 (1)	16	28	18	73
2	7	1 (1)	6	12	10	27
3	13	4 (1)	10	22	14	59
Totals	27	11 (3)	32	62	42	159

(1) = number of sub-conjugates in which different pure line types were found: delayed reorganization, delayed change in nuclear constitution

nearly all possible combinations were found within sister conjugates: (except that

8. Crossing same mating types, $\text{II} \times \text{V}$ from F₂ individuals, on examples -- yielded very different mating types. And, different allelic II or V just different spectra of types

Table 3 of paper:

Cross	Temperature	mating types of conjugates from $\text{II} \times \text{V}$								Total
		I	II	III	IV	V	VI	VII	Selfing culture	
4	16°C	2	5	4	0	5	13	0	9	38
	26°C	12	3	4	0	3	4	0	9	35
5	16°C	0	5	4	11	2	0	2	14	38
	26°C	0	11	1	9	2	0	4	16	43

Interpretation (mine)

Results suggest an unstable locus determining substance responsible for mating type. In *P. aurelia*, time of change at locus occurs, mainly, at first division of macronucleus in sister conjugants, ^{due to two parental types} In *Tetrahymena*, time of change has greater spread, going into sub-caryonuclear divisions but mainly at first division of macronucleus. Here, type of change can give a number of new mating types, not found in parents reaction types.

Time and type of change in nuclear constitution illustrated.

Related to mitotic event.

Auto-mutagenesis in ^{the} mushrooms, *Sclizophyllum commune*

John R. Roper, Univ. of Chicago. Science 118 pg 574, 1953 (AA).

1. Tetrapolar sexuality; obligate crossmating imposed and controlled by incompatibility factors at two loci, A + B.
2. Each fruit produces progeny of 4 distinct mating types
3. Fertile matings occur between strains of these diverse mating types only in those combinations which yield the double heterozygote.
4. In other combinations having A + B factors in common, heterokaryons of a unique type are formed.
5. The common - A heterokaryon, which is stable and capable of indefinite vegetative growth, induces mutation in its component nuclei in frequencies up to 10^6 times the ~~occurs~~ spontaneous rate for the same mutations in ~~these~~ ^{these} ~~large~~ ^{large} strains.
6. Mutants belong to a limited number of distinct morphological types (about 10) each differing from the wild by a single, characteristic, altered locus.
7. Age of heterokaryon + genetic history of its component strains determine mutation frequency and, to a lesser extent, the type of mutation induced.
8. ^{Auto-induced} Mutagen agent's suspected that preferentially affects a limited number of loci in a regular and predictable manner.

Hinton, T., P. T. Yuen & A. T. Evans

Changing the gene order and number in natural populations

Isolation 6: 19-28

1.

1. 13 x chr. mutant stocks examined. derived from by of Dues.
2. Each carries inversion from c.o. studies
3. Above in the stock with frequency of 4.8% of tested lethal-bearing chromosomes.
4. Kept in stock by balancing with A-49.
5. Crossed to normal & salivary examined.

Breaks.	M (1) h1	Breaks	
	1	12 E	20
	" 2	1 F	20
	" 3	4 D	20
same locus	" 4	4 C	20
	" 5	1 F	20
same locus	" 6	4 C	20
	" 7	12 E	20
same locus	" 8	3 C	20
	" 9	8 F	20
	" 10	4 E 2, 3	8 A, 1, 2
	" 12	1 C 3	20
	" 13	4 E	20

Transposition (h1 11) Break. at 5C, 7E, 20. Region between 5C and 7E inserted up to 20.

nearly 1/2 of breaks occur in region 4.

Regions involved coincide with positions of hetero chromatin given by Kaufmann

Break positions in 20 not determined cytologically. Cytosol studies suggest separation of 1/2 of breaks in 20.

Species crosses - attraction under complement

I. Many examples - change in gene action,

appearance of migration

Changes in chemical constituents - odors

but ^{same} - Change in set - Median
salivary glands. Specimen A x B.

Some cells = Hybrids

" " = all of A

" " = Part of A + B.

Median - elementary change in set in

f.c. $A + B \times A = B$ also
elementary contrast

Ds type behavior - Besides many.

1. Hyacinth - Darlington. Particularly obvious - breaks late in development at specific regions of a specific chromosome.
2. Secura - Metz. Strain showing break at specific region of specific chromosome in strain of Secura. Genetically controlled.
3. Duffus - Break at specific region of certain chromosomes during spermatogenic divisions.
4. Vicia - Levan - Break at specific regions ^{m.o.} in cells in early development - same environment.
5. Oenothera - Catcheside. Evidence strongly suggests Ds type behavior in chromosomes arising from trisecation. Breaks at or close to heterochromatin.

Differential segregation - genetic, ^{expression} non-Mendelian. During meiosis ^{to gametes}

1. Paramutation - mating type reaction - Two alleles ^{mutant} differ following this division. At certain time in development.
2. Tetrahymena - ^{in vivo} Paramutation not following Mendel's law.
3. Case in Glomella where "mutation" is genetically controlled in the development of the neoplasms. At certain time ^{well}.

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4. Control of patterns - why pigment found at certain places
 5. Wing of *Cybertria* - differential mitosis required.

Differences in number in different cells of an organism: Morphological Studies

1. Zygote = $2n$ = 2 sets of chromosomes.

- a). Increasing number of sets ^① - definite degrees in maturity cells of tissues
Widespread phenomenon ^② Number - very commonly: Polytens
 strands remain together.

Differ - strands

separated

Combination of both

b). Decreasing sets of chromosomes - at particular time in development

- ① Frog - Tail Mesenchyme cells disappear in
 Epidermis = apoptosis

② Sea urchin - Somatic reduction in division of certain,
 clearly differentiated cells of 16 cell stage of embryo - the mesenchyme.

all 4 mesenchyme nuclei = $2n$ (36)

during their division = reduction in molecular masses. (18)

③ Cells (marginal)

2. Differences in different cells of organism with regard to certain

Characteristics of the complement: Science

[the organism]
 or

a) Maize - B-type chromosomes. [the continuous series]

①. Plants & Animals - many cases like that of both Maize & Maize.

3. Differences in behavior of sets of chromosomes ^{within} in a nucleus - Report differences in function.

one set condensed = become dense in sperm (white) -

///
/

one set extended.

4. Differences in behavior of individual chromosomes in complement at different times & in different cells of lion
a) White - salivary; Tulland - salivary.

b). Some cells  others

Different

same

5. Differences in behavior of individual parts of chromosomes.

a) Large

region

cells
↓ or ↓

b). Chromocenter region - Drosophila

Somatic cells - mouse

Salivary.

6) Smallest segment known - the band in salivary glands - Pavan.

Rhyndiscus:

1) all eggs laid within 45 min

2) larvae = all of 1 set

3) " develop all at same time 42 days to pupae.

4) Extraordinary salivary gland type cluster in several of the lines

5) Can follow differences between individual bands at any one time.

6) Shown: certain bands in particular lines of all larvae undergo same type of change during development + some sequences of changes. Reflects differential action at certain times in development.

Pavan interpretation of some changes: increase in DNA, or of gene components, differentially in a particular band. Always same bands, in same times at same time in development.

March 4th.

I. Review of previous discussion.

1. Purpose - to show that other systems present that control genetic expressions besides the Ds-A + G1-DT system, which are much older.

a) To show that some basic similarities are present - that controlling systems ^{are present} + showing the clear-cut somatic segregation & Mendelian segregation that give the same change in gene expression. Also, that initial evidence supports transposition of the controlling factors.

a) Such transposons have also been proposed to account for genetic change in expression of the var. ^{of P. l.} ^{in many} + another case of variegated pale green by Pelton.

II

Since these controlling systems ^{have} ~~make~~ them selves evident in cells whose nuclear composition has been altered, we infer that they were present in the nucleus before the alteration occurred. ^{Therefore, as} ^{some members -- controlling gene expression down to at specific times in specific cells.}
How can we account for their appearance:

If we take the view that they are present in the nucleus before alteration, then they must be functioning in normal nuclei -- (1) They seem to control gene actions at certain stages of development. (2) That the somatic segregation ^{is} occurring during normal development. (3) That the cells in which such segregation occur do not give rise to the same cells but to cells which will be differentiating.

in one "set line of determined course. (4). Differentiation & budding
vegetating will not arise from cells which have lost competence by the
regeneration process but can arise only from specific cells within the
tissue, which have not been altered by somatic segregation.

The reasons we have been able to see this process is because the
regeneration process has been put - "out of phase" by the alterations
occurring to the heterochromatin elements by the L.P. 6. cycle.

We can therefore examine this process because it is "out of
phase" and determine the factors involved and how they operate.

This would require that, the ^{cytotax} nucleus of differentiated cells of
an organism would vary genetically. They would not be ^{in normal development} identical
to one another genetically. It would require controlled types of
somatic segregation at species dimensionsally controlled. It would
suggest that chromosomal alterations - translocations etc - might
accompany ^{some of} these changes. What evidence do we have that

such events occur in normal development? There is a vast

amount of ^{embryonic} evidence pointing in this direction. Some representative
examples will be chosen.

III. The "dedifferentiation" of differentiated plant cells:

1. Cells not not fully differentiated usually would not undergo a change
2. These forced to undergo a change by chemical treatment - (This treatment does not induce chemical alteration in the already cells)
3. The chemical compound -- chemical alteration, present in water, was present in these cells. The polyphenol cells arising from protoplasts:

$$\gg \pi = (($$

$$= "$$

The anaphase. Bridges & fragments. Two homologues. $\left[\begin{array}{c} \text{---} \\ \text{---} \end{array} \right] \subset \left[\begin{array}{c} \text{---} \\ \text{---} \end{array} \right]$
 Same alteration in each homologue. Expresses controlled event at same position in each homologue.

4. Such cells not capable of developing a new organism because of many alterations present that are exposed at anaphase.
5. Conclude that the chemical alterations are of a controlled nature - Association of particular compounds of the nucleus in particular cells.

IV. The controlled segregation of nuclear components during normal development. These occur on all levels - from chromosomes sets to components of chromosomes.

1. Best analyzed case - *Sacchara* Many other with same general pattern.
2. The B-type chromosome in plants - Many cases - Examples B-type Chr.
3. Genetic non-equivalency - Differentiated segregates
Paramecium malingtys
Glomerella " "

V. Non-equivalency of chromosomal components in normal development - all levels. from sets, to ^{units} chromosomes, to ^{parts} parts of chromosomes, to molecules within chromosomes.

See sheet.